

Patients With IgA Nephropathy Have Altered Levels of Immunomodulatory C19 Steroids. Glucocorticoid Therapy With Addition of Adrenal Androgens May Be the Choice

I. ŠTERZL¹, M. HILL¹, L. STÁRKA¹, M. VELÍKOVÁ¹, R. KANČEVA¹, J. JEMELKOVÁ²,
L. CERNEKOVÁ², P. KOSZTYU², J. ZADRAŽIL², K. MATOUŠOVIC³, K. VONDRAK³,
M. RAŠKA²

¹Institute of Endocrinology, Prague, Czech Republic, ²Palacky University and University Hospital Olomouc, Olomouc, Czech Republic, ³Second Faculty of Medicine, Charles University and Motol University Hospital, Prague, Czech Republic

Received July 11, 2017

Accepted July 16, 2017

Summary

Glucocorticoid (GC) therapy is one of the methods of choices for treatment of autoimmune diseases (ADs). In addition, adrenal androgens are known as immunoprotective GC-antagonists. Adrenal steroids preferentially influence the Th1-components over the Th2 ones. We investigated steroid metabolome (using gas chromatography-mass spectrometry) in healthy controls (H), GC-untreated patients with ADs different from IgA nephropathy (U), GC-treated patients with ADs different from IgA nephropathy (T) and in patients with IgA nephropathy (IgAN), which were monitored on the beginning (N0), after one week (N1) and after one month (N2) of prednisolone therapy (60 mg of prednisolone/day/m² of body surface). Between-group differences were assessed by one-way ANOVA, while the changes during the therapy were evaluated by repeated measures ANOVA. The ANOVA testing was followed by Duncan's multiple comparisons. IgAN patients and patients with other ADs exhibited lack of adrenal androgens due to attenuated activity of adrenal *zona reticularis* (ZR). Androgen levels including their 7α-, 7β-, and 16α-hydroxy-metabolites were further restrained by GC-therapy. Based on these results and data from the literature, we addressed the question, whether a combination of GCs with Δ⁵-steroids or their more stable synthetic derivatives may be optimal for the treatment of antibodies-mediated ADs.

Key words

Glucocorticoid therapy • Adrenal androgens • Immunomodulatory steroids • Gas chromatography-mass spectrometry • Steroid metabolome

Corresponding author

M. Hill, Institute of Endocrinology, Národní třída 8, 116 94 Prague 1, Czech Republic. E-mail: mhill@endo.cz

Introduction

IgA nephropathy (IgAN) is the most frequent primary glomerulonephritis worldwide. In this autoimmune disease, aberrantly O-glycosylated IgA1 hinge region serves as an antigen recognized by anti-glycan antibodies. This stimulates a formation of nephritogenic immune complexes (Mestecky *et al.* 2013), the accumulation of which in the kidney mesangium induces a proliferation of mesangial cells, expansion of extracellular matrix proteins and renal injury, leading to an end-stage renal failure in 20–40 % of patients. Progressive IgAN patients may be treated with glucocorticoids (GCs) (Lai *et al.* 2016, Matousovic *et al.* 2015). However, this therapy in IgAN is questionable (Lv *et al.* 2012, Rauen *et al.* 2015, Tesar *et al.* 2015). Although GC-administration represents the most powerful therapy for rapid attenuation of inflammatory response, longtime GC-treatment is accompanied by serious complications such as disruption of hypothalamo-pituitary-adrenal (HPA) axis and inhibition of gonadal activity (Mastorakos *et al.* 2006, Rengarajan and Balasubramanian 2008). As adrenal androgens are GC-antagonists, we evaluated their levels in groups of healthy controls (H), GC-untreated (U) and GC-treated

(T) patients with other autoimmune diseases (ADs), and in IgAN patients before prednisolone treatment (N0), and after one week (N1) and one month (N2) of prednisolone therapy. Since natural $7\alpha/\beta$ - and 16α -hydroxy-steroids are immunomodulatory, anti-inflammatory and immunoprotective (Ahlem *et al.* 2011a, Ahlem *et al.* 2011b, Auci *et al.* 2009, Conrad *et al.* 2010, Hennebert *et al.* 2007, Le Mee *et al.* 2008, Loria 2002, Pettersson *et al.* 2010, Reading *et al.* 2012, Tang *et al.* 2006) we focused on these substances.

Methods

Subjects

The study enrolled 23 women and 43 men. The study groups consisted of healthy controls (H, n=10), untreated controls without IgAN but suffering from other ADs (U, n=10), GC treated non-IgAN controls suffering from other ADs (T, n=9), and patients with IgAN (n=14) who were monitored before prednisolone treatment (N0), after one week (N1) and one month (N2) of prednisolone therapy. The U-group enrolled patients with idiopathic membranous glomerulonephritis, nephrotic syndrome, membranous glomerulonephritis, glomerulopathy, and suspenzia chronic mesangioproliferative glomerulonephritis. The T-group consisted of patients with rheumatoid arthritis, systemic lupus erythematosus, ankylosing spondylitis, systemic impairment of connective tissues, polymyalgia rheumatica and giant cell arteritis, membranous nephropathy, complete nephrotic syndrome and glomerular and segmental glomerulosclerosis. Prednisolone dose was 60 mg of prednisolone/day/m² of body surface. The patients have been recruited from the Nephrological Outpatient Department (NOPD) of the Motol University Hospital and from the neighboring NOPDs in Prague and NOPD of Palacky University Olomouc. No samples from the female subjects were collected in the luteal menstrual phase. The mean age of the participants was 48.4±15.1 years (mean ± SD) and the age differences between the study groups were insignificant.

The Ethics Committee of the Institute of Endocrinology in Prague approved the protocol of the study, and written informed consent was obtained from all participants.

Analytical methods

The circulating levels of steroids and their polar conjugates were measured using a previously described

gas chromatographic-mass spectrometric method (Hill *et al.* 2010).

Statistical analysis

The between-group differences were evaluated using one-way ANOVA. Changes in steroid levels were assessed using repeated measures ANOVA. The ANOVA testing was followed by Duncan's multiple comparisons (Duncan 1955). The original data were transformed by power transformations to attain Gaussian data distribution and constant variance. Statgraphics Centurion, version XV statistical software from Manugistics, (Herndon, MA, USA) was used for the calculations.

Results

Steroid levels

Most steroids showed significant between-group differences (Table 1) and there was a trend of decreasing in steroid levels during the prednisolone therapy (Table 2). Estradiol was higher in the IgAN patients irrespectively of GC-treatment.

Shifted balance between adrenal C21 and C19- Δ^5 -steroids towards the C21 steroids in IgAN patients

Dehydroepiandrosterone (DHEA) to pregnenolone ratio (Fig. 1A) was higher in the H-group than in the other groups while the dehydroepiandrosterone sulfate (DHEAS) to pregnenolone sulfate ratio did not significantly differ between the H and N0-groups (Fig. 1B). Both ratios (reflecting CYP17A1 – lyase step) decreased during the GC-treatment of IgAN patients. The values and statistics for repeated measures ANOVA in the DHEA/pregnenolone ratio were F=26.51, p<0.001: N0=5.66 (4.72, 6.73 – mean with 95 % confidence limits), N1=2.7 (2.14, 3.36), N2=2.06 (1.57, 2.64), N0-N1, N0-N2 (significant differences between stages N0 and N1 and between stages N0 and N2). For the DHEAS/pregnenolone sulfate ratio they were F=4.02, p=0.031: N0=15.2 (12.3, 18.6), N1=11.9 (9.69, 14.5), N2=10.1 (8.22, 12.4), N0-N2.

Increased conversion of DHEA to androstenedione reflecting type 2 3 β -hydroxysteroid dehydrogenase (HSD3B2) activity in IgAN patients

Androstenedione/DHEA (Fig. 1C) and androstenedione/DHEAS (Fig. 1D) ratios reflecting type 2 3 β -hydroxysteroid dehydrogenase (HSD3B2) activity were higher in the GC-treated groups. The values

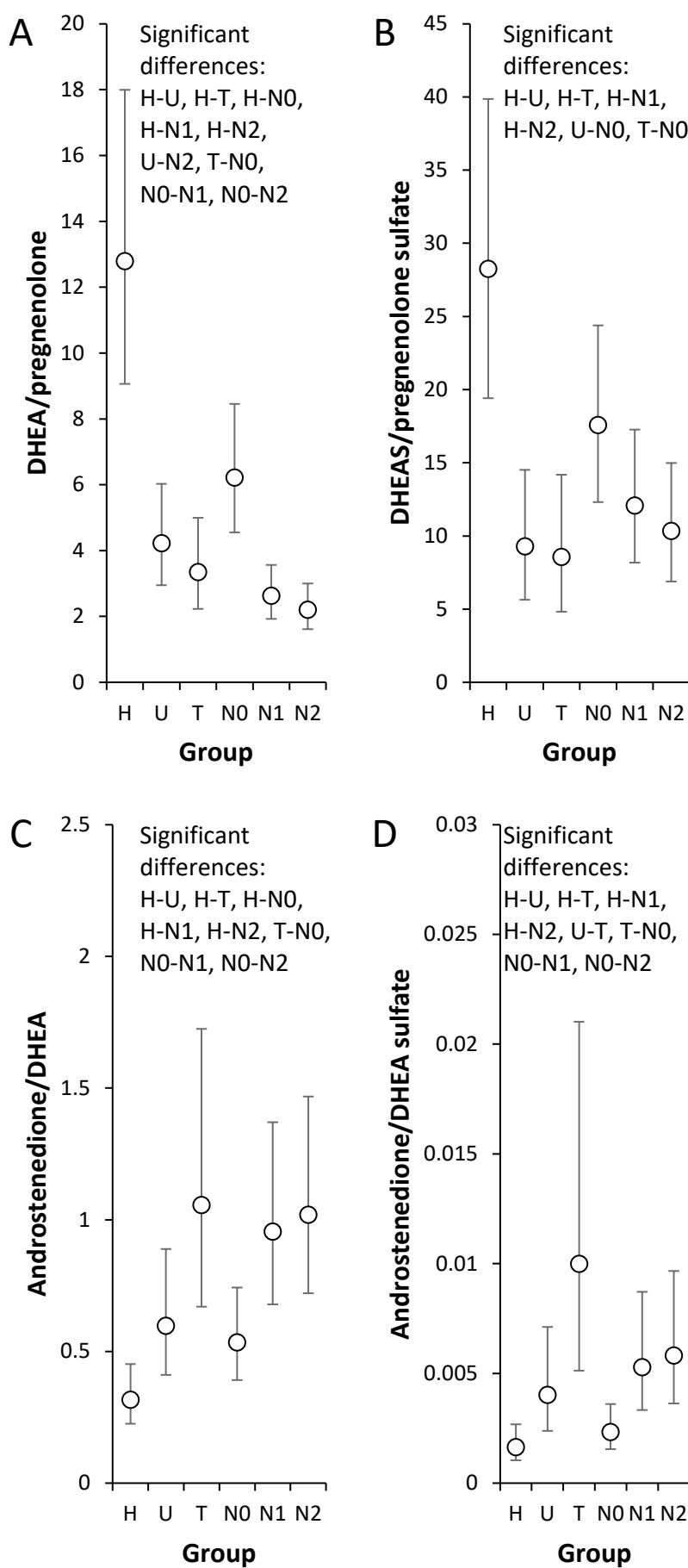


Fig. 1. Product to precursor ratios (PPRs) in groups of healthy controls (H), glucocorticoid-untreated patients with autoimmune diseases (ADs) different from IgA nephropathy (IgAN) (U), GC-treated patients with non-IgAN ADs (T) and in IgAN patients on the beginning of prednisolone treatment (N0), after one week (N1) and after one month (N2) of the therapy. The circles with error bars represent re-transformed means with their 95 % confidence intervals (for details see section Statistical analysis). The embedded tables show all significant between-group differences ($p<0.05$) as evaluated by Duncan's multiple comparisons. The symbols separated by hyphen represent significant between-group differences ($p<0.05$). Panels **A** and **B** show the PPRs reflecting a balance between adrenal C21 and C19- Δ^5 -steroids, while panels **C** and **D** show the PPRs reflecting the activity of type 2 17 β -hydroxysteroid dehydrogenase (HSD17B2).

Table 1. Levels of Δ^5 -steroids, their metabolites and estradiol in healthy controls (H), corticoid-untreated (U) and corticoid-treated patients with autoimmune diseases different from IgA nephropathy (T) and patients with IgA nephropathy on the beginning of prednisolone therapy (N0), on one week (N1) and one month (N2) of prednisolone therapy.

Steroid [nmol/l]	H	U	T	N0	N1	N2
<i>Pregnенолоне</i>	0.884 (0.568, 1.47)	0.91 (0.582, 1.52)	0.666 (0.423, 1.13) <i>F=0.64, p=0.670</i>	1.07 (0.72, 1.69)	0.846 (0.583, 1.29)	0.71 (0.498, 1.06)
<i>Pregnенолоне сулфат</i>	82.8 (51.6, 132)	72 (44.7, 115) <i>F=5.32, p<0.001, H-T, H-NI, H-N2, U-T, U-NI, U-N2, T-NI, N0-NI, N0-N2</i>	29.9 (17.2, 51.3) <i>F=6.84, p<0.001, H-U, H-T, H-NI, H-N2, N0-NI, N0-N2</i>	87.2 (58.6, 129) <i>F=9.35, p<0.001, H-T, H-NI, H-N2, U-T, U-NI, U-N2, T-NI, N0-NI, N0-N2</i>	35.8 (23.7, 53.7) <i>F=6.84, p<0.001, H-U, H-T, H-NI, H-N2, N0-NI, N0-N2</i>	30.8 (20.3, 46.3) <i>F=1.1 (0.819, 1.48)</i>
<i>20α-Dihydro pregnenolone</i>	2.64 (1.85, 3.8)	1.53 (1.08, 2.19) <i>F=9.35, p<0.001, H-T, H-NI, H-N2, U-T, U-NI, U-N2, T-NI, N0-NI, N0-N2</i>	1.28 (0.871, 1.91) <i>F=3.92, p=0.004, H-T, U-NI, T-NI, T-N2</i>	1.86 (1.38, 2.52) <i>F=3.92, p=0.004, H-T, U-NI, T-NI, T-N2</i>	1.1 (0.819, 1.48) <i>F=3.92, p=0.004, H-T, U-NI, T-NI, T-N2</i>	0.962 (0.719, 1.29) <i>F=1.1 (0.819, 1.48)</i>
<i>20α-Dihydro pregnenolone sulfate</i>	421 (240, 688)	348 (193, 582) <i>F=9.35, p<0.001, H-T, H-NI, H-N2, U-T, U-NI, U-N2, T-NI, N0-NI, N0-N2</i>	144 (61.2, 291) <i>F=9.35, p<0.001, H-T, H-NI, H-N2, U-T, U-NI, U-N2, T-NI, N0-NI, N0-N2</i>	336 (204, 524) <i>F=9.35, p<0.001, H-T, H-NI, H-N2, U-T, U-NI, U-N2, T-NI, N0-NI, N0-N2</i>	117 (60, 207) <i>F=9.35, p<0.001, H-T, H-NI, H-N2, U-T, U-NI, U-N2, T-NI, N0-NI, N0-N2</i>	113 (57.5, 201) <i>F=1.1 (0.819, 1.48)</i>
<i>16α-Hydroxy pregnenolone</i>	0.256 (0.129, 0.508)	0.172 (0.0867, 0.34) <i>F=3.92, p=0.004, H-T, U-NI, T-NI, T-N2</i>	0.077 (0.036, 0.164) <i>F=3.92, p=0.004, H-T, U-NI, T-NI, T-N2</i>	0.56 (0.314, 1) <i>F=3.92, p=0.004, H-T, U-NI, T-NI, T-N2</i>	0.325 (0.182, 0.581) <i>F=3.92, p=0.004, H-T, U-NI, T-NI, T-N2</i>	0.218 (0.123, 0.39) <i>F=1.1 (0.819, 1.48)</i>
<i>Dehydroepiandrosterone (DHEA)</i>	11.9 (6.88, 20.3)	4.42 (2.5, 7.7) <i>F=7.71, p<0.001, H-U, H-T, H-NI, H-N2, U-N2, T-NI, N0-NI, N0-N2</i>	2.38 (1.24, 4.49) <i>F=7.71, p<0.001, H-U, H-T, H-NI, H-N2, U-N2, T-NI, N0-NI, N0-N2</i>	6.43 (4.01, 10.2) <i>F=7.71, p<0.001, H-U, H-T, H-NI, H-N2, U-N2, T-NI, N0-NI, N0-N2</i>	2.35 (1.44, 3.81) <i>F=7.71, p<0.001, H-U, H-T, H-NI, H-N2, U-N2, T-NI, N0-NI, N0-N2</i>	1.75 (1.07, 2.85) <i>F=1.1 (0.819, 1.48)</i>
<i>DHEA sulfate</i>	2310 (1270, 3990)	749 (365, 1430) <i>F=7.26, p<0.001, H-U, H-T, H-NI, H-N2, T-NI, N0-NI, N0-N2</i>	276 (106, 635) <i>F=3.55, p=0.007, H-U, H-T, H-NI, H-N2, T-NI, N0-NI, N0-N2</i>	1520 (886, 2490) <i>F=3.55, p=0.007, H-U, H-T, H-NI, H-N2, T-NI, N0-NI, N0-N2</i>	429 (222, 780) <i>F=3.55, p=0.007, H-U, H-T, H-NI, H-N2, T-NI, N0-NI, N0-N2</i>	327 (164, 610) <i>F=1.1 (0.819, 1.48)</i>
<i>7α-Hydroxy-DHEA</i>	0.643 (0.37, 1.09)	0.349 (0.195, 0.608) <i>F=2.90, p=0.021, H-T, T-NI, T-N2</i>	0.161 (0.08, 0.313) <i>F=2.90, p=0.021, H-T, T-NI, T-N2</i>	0.566 (0.354, 0.889) <i>F=2.90, p=0.021, H-T, T-NI, T-N2</i>	0.234 (0.138, 0.387) <i>F=2.90, p=0.021, H-T, T-NI, T-N2</i>	0.285 (0.173, 0.461) <i>F=1.1 (0.819, 1.48)</i>
<i>7β-Hydroxy-DHEA</i>	0.36 (0.217, 0.612)	0.208 (0.128, 0.346) <i>F=1.75, p<0.136, H-T, H-N2</i>	0.116 (0.0665, 0.207) <i>F=1.75, p<0.136, H-T, H-N2</i>	0.403 (0.261, 0.632) <i>F=1.75, p<0.136, H-T, H-N2</i>	0.278 (0.179, 0.438) <i>F=1.75, p<0.136, H-T, H-N2</i>	0.228 (0.15, 0.351) <i>F=1.1 (0.819, 1.48)</i>
<i>Androstenediol</i>	1.57 (0.868, 2.66)	0.865 (0.433, 1.57) <i>F=2.42, p<0.045, H-T, T-NI</i>	0.577 (0.228, 1.24) <i>F=2.42, p<0.045, H-T, T-NI</i>	1.21 (0.714, 1.95) <i>F=2.42, p<0.045, H-T, T-NI</i>	0.712 (0.389, 1.21) <i>F=2.42, p<0.045, H-T, T-NI</i>	0.61 (0.326, 1.06) <i>F=1.1 (0.819, 1.48)</i>
<i>Androstenediol sulfate</i>	612 (271, 1230)	190 (70.3, 435) <i>F=1.91, p=0.105, H-N2</i>	120 (34.3, 324) <i>F=1.91, p=0.105, H-N2</i>	52.8 (272, 948) <i>F=1.91, p=0.105, H-N2</i>	332 (160, 630) <i>F=1.91, p=0.105, H-N2</i>	193 (84.7, 391) <i>F=1.1 (0.819, 1.48)</i>
<i>5-Androsten-3β,7α,17β-triol</i>	0.065 (0.030, 0.153)	0.032 (0.016, 0.069) <i>F=3.35, p=0.010, U-NI, T-NI</i>	0.025 (0.012, 0.056) <i>F=3.35, p=0.010, U-NI, T-NI</i>	0.177 (0.084, 0.415) <i>F=3.35, p=0.010, U-NI, T-NI</i>	0.084 (0.043, 0.177) <i>F=3.35, p=0.010, U-NI, T-NI</i>	0.068 (0.036, 0.142) <i>F=1.1 (0.819, 1.48)</i>
<i>5-Androsten-3β,7β,17β-triol</i>	0.058 (0.026, 0.143)	0.023 (0.012, 0.050) <i>F=3.48, p=0.008, H-T, U-NI, T-NI, T-N2</i>	0.016 (0.008, 0.037) <i>F=3.48, p=0.008, H-T, U-NI, T-NI, T-N2</i>	0.124 (0.058, 0.299) <i>F=3.48, p=0.008, H-T, U-NI, T-NI, T-N2</i>	0.053 (0.027, 0.112) <i>F=3.48, p=0.008, H-T, U-NI, T-NI, T-N2</i>	0.037 (0.020, 0.075) <i>F=1.1 (0.819, 1.48)</i>
<i>Androstenedione</i>	3.79 (2.47, 5.77)	2.74 (1.77, 4.2) <i>F=3.20, p=0.014, U-NI, U-N2, T-NI, T-N2</i>	2.57 (1.57, 4.16) <i>F=3.20, p=0.014, U-NI, U-N2, T-NI, T-N2</i>	3.54 (2.46, 5.05) <i>F=3.20, p=0.014, U-NI, U-N2, T-NI, T-N2</i>	2.29 (1.58, 3.3) <i>F=3.20, p=0.014, U-NI, U-N2, T-NI, T-N2</i>	1.86 (1.28, 2.7) <i>F=1.1 (0.819, 1.48)</i>
<i>Estradiol</i>	0.046 (0.021, 0.10)	0.027 (0.013, 0.059) <i>F=3.20, p=0.014, U-NI, U-N2, T-NI, T-N2</i>	0.030 (0.013, 0.071) <i>F=3.20, p=0.014, U-NI, U-N2, T-NI, T-N2</i>	0.143 (0.064, 0.333) <i>F=3.20, p=0.014, U-NI, U-N2, T-NI, T-N2</i>	0.108 (0.048, 0.248) <i>F=3.20, p=0.014, U-NI, U-N2, T-NI, T-N2</i>	0.106 (0.048, 0.244) <i>F=1.1 (0.819, 1.48)</i>

The differences were evaluated by one-way ANOVA after Box-Cox transformations of original data for attainment of Gaussian distribution and constant variance. The ANOVA testing was followed by Duncan's multiple comparisons. F and p represent F-statistics and their p-values, respectively. The steroid levels are shown as retransformed means with their 95 % confidence intervals. F represents Fisher's statistics in ANOVA testing and p symbolizes its significance level. The symbols separated by hyphen represent significant between-group differences ($p < 0.05$).

Table 2. Levels of Δ^5 -steroids and their metabolites in patients with IgA nephropathy on the beginning of prednisolone therapy (N0), on one week (N1), and one month (N2) of prednisolone therapy.

Steroid [nmol/l]	Mean (lower 95 % confidence limit, upper 95 % confidence limit)			Factor Stage*
	N0	N1	N2	
Pregnolone	1.05 (0.81, 1.39)	0.833 (0.658, 1.08)	0.691 (0.555, 0.878)	F=2.96, p=0.069, N0-N2
Pregnolone sulfate	87.2 (68.3, 111)	35.8 (27.8, 46)	30.8 (23.9, 39.6)	F=21.8, p<0.001, N0-N1, N0-N2
20 α -Dihydro pregnenolone	1.83 (1.47, 2.29)	1.07 (0.877, 1.32)	0.949 (0.779, 1.16)	F=11.5, p<0.001, N0-N1, N0-N2
20 α -Dihydro pregnenolone sulfate	299 (231, 388)	107 (82.8, 139)	101 (77.7, 130)	F=23.6, p<0.001, N0-N1, N0-N2
16 α -Hydroxy pregnenolone	0.552 (0.409, 0.748)	0.321 (0.239, 0.434)	0.215 (0.16, 0.289)	F=10.6, p<0.001, N0-N1, N0-N2
Dihydroepiandrosterone (DHEA)	6.48 (5.07, 8.25)	2.37 (1.82, 3.08)	1.77 (1.35, 2.32)	F=29.9, p<0.001, N0-N1, N0-N2
DHEA sulfate	1280 (937, 1790)	396 (313, 510)	266 (214, 334)	F=39.0, p<0.001, N0-N1, N0-N2, N1-N2
7 α -Hydroxy-DHEA	0.555 (0.364, 0.849)	0.28 (0.184, 0.426)	0.267 (0.176, 0.406)	F=4.02, p=0.003, N0-N1, N0-N2
7 β -Hydroxy-DHEA	0.414 (0.306, 0.556)	0.279 (0.198, 0.392)	0.246 (0.18, 0.334)	F=3.34, p=0.052, N0-N2
Androstenediol	1.46 (1.2, 1.74)	0.779 (0.603, 0.978)	0.688 (0.523, 0.875)	F=14.6, p<0.001, N0-N1, N0-N2
Androstenediol sulfate	435 (352, 539)	280 (227, 346)	178 (144, 222)	F=18.2, p<0.001, N0-N1, N0-N2, N1-N2
5-Androsten-3 β ,7 α ,17 β -triol	0.182 (0.128, 0.265)	0.086 (0.062, 0.121)	0.071 (0.052, 0.099)	F=8.90, p=0.001, N0-N1, N0-N2
5-Androsten-3 β ,7 β ,17 β -triol	0.145 (0.098, 0.216)	0.060 (0.041, 0.088)	0.045 (0.031, 0.066)	F=10.6, p<0.001, N0-N1, N0-N2
Androstenedione	3.72 (3.02, 4.51)	2.83 (2.19, 3.55)	2.27 (1.74, 2.89)	F=5.19, p=0.013, N0-N2
Estradiol	0.187 (0.131, 0.244)	0.203 (0.15, 0.255)	0.13 (0.0689, 0.192)	F=1.92, p=0.181

*The changes in steroid levels were evaluated by repeated measures ANOVA consisting of factors Stage and Subject after Box-Cox transformations of original data for attainment of Gaussian distribution and constant variance. For explanation of remaining symbols, see Table 1. The ANOVA testing was followed by Duncan's multiple comparisons. F and p represent F-statistics and their p-values, respectively. The steroid levels are shown as retransformed means with their 95 % confidence intervals. F represents Fisher's statistics in ANOVA testing and p symbolizes its significance level. The symbols separated by hyphen represent significant between-group differences ($p<0.05$).

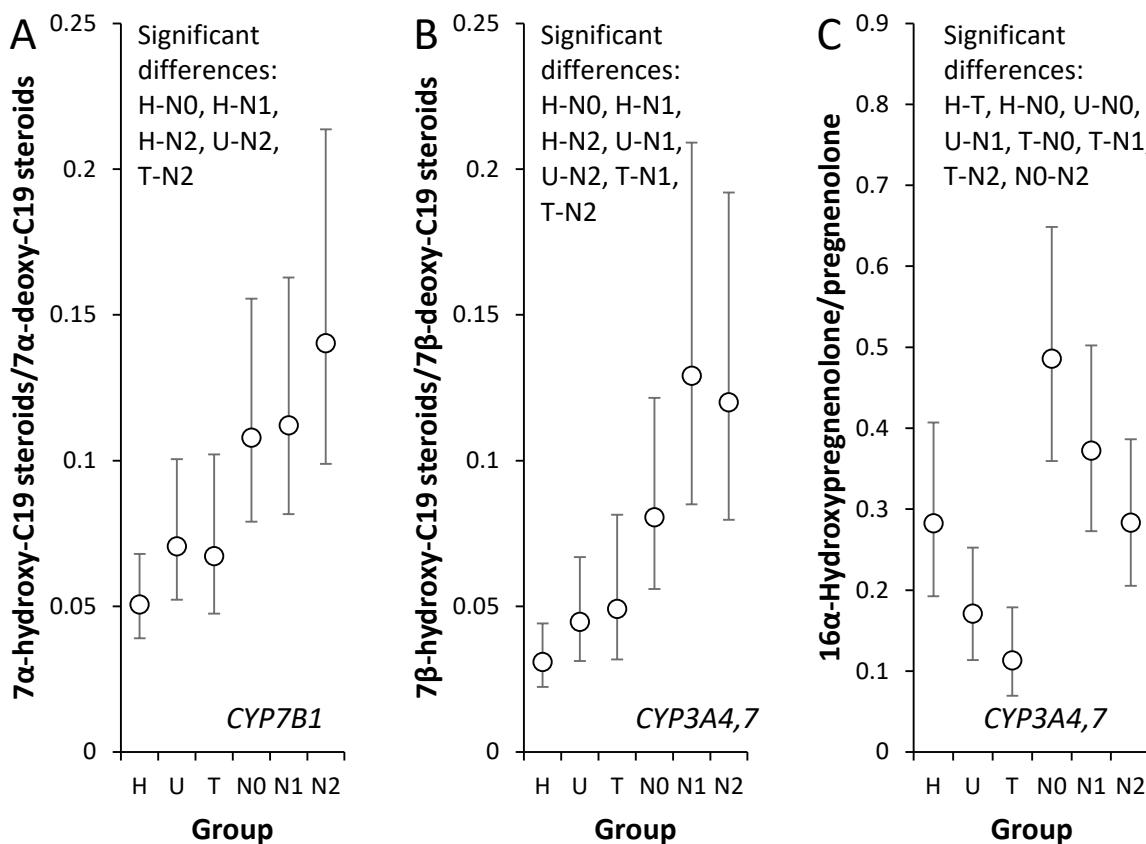


Fig. 2. Product to precursor ratios (PPRs) in groups of healthy controls (H), glucocorticoid-untreated patients with autoimmune diseases (AD) different from IgA nephropathy (IgAN) (U), GC-treated patients with non-IgAN AD (T) and in IgAN patients on the beginning of prednisolone treatment (N0), after one week (N1) and after one month (N2) of the therapy. The text in italics illustrates the key enzymes, which may be reflected by the PPRs. For other drawings and symbols, see Figure 1. Panel **A** shows the PPRs reflecting a conversion of DHEA and androstenediol to their immunomodulatory 7α -hydroxy-metabolites. Panel **B** illustrates the PPRs reflecting a conversion of DHEA and androstenediol to their immunomodulatory 7β -hydroxy-metabolites. Panel **C** illustrates the PPRs reflecting a conversion of pregnenolone to 16α -hydroxypregnenolone.

and statistics for repeated measures ANOVA for the androstenedione/DHEA ratio were $F=7.23$, $p=0.003$: N0=0.561 (0.422, 0.738), N1=0.983 (0.755, 1.27), N2=1.07 (0.824, 1.38), N0-N1, N0-N2. For the androstenedione/DHEAS ratio they were $F=17.61$, $p<0.001$: N0=0.0028 (0.0021, 0.0037), N1=0.0066 (0.0052, 0.0081), N2=0.0067 (0.0054, 0.0081), N0-N1, N0-N2. The values in the H-groups were lower in comparison with the other groups for the androstenedione/DHEA ratio and similar results were found for the androstenedione/DHEAS ratio except for insignificant difference between the H and N0-groups.

Changing 7α -, 7β - and 16α -hydroxylation of Δ^5 -steroids in IgAN patients during the prednisolone treatment

Figure 2A shows the product to precursor ratio based on the levels of 7α -hydroxy-DHEA (DHEA 7α), 5-androstene-3 β , 7α ,17 β -triol (AT 7α), DHEA and androstenediol. This $(\text{DHEA}7\alpha+\text{AT}7\alpha)/(\text{DHEA}+\text{androstenediol})$ ratio exemplifies a conversion of DHEA

and androstenediol to their 7α -hydroxy-metabolites. This ratio was elevated in IgAN groups and showed an increase at the second stage of GC-therapy of IgAN patients ($F=5.14$, $p=0.014$: N0=0.101 (0.085, 0.122), N1=0.094 (0.079, 0.114), N2=0.139 (0.115, 0.173), N0-N2, N1-N2).

Figure 2B shows the product to precursor ratio based on the levels of 7β -hydroxy-DHEA (DHEA 7β), 5-androstene-3 β , 7β ,17 β -triol (AT 7β), DHEA and androstenediol. This $(\text{DHEA}7\beta+\text{AT}7\beta)/(\text{DHEA}+\text{androstenediol})$ ratio illustrating a conversion of DHEA and androstenediol to their immunomodulatory 7β -hydroxy-metabolites exhibited a similar picture as in the case of 7α -hydroxylation, except the transient increase during the GC-treatment ($F=2.99$, $p=0.068$: N0=0.079 (0.061, 0.105), N1=0.124 (0.0914, 0.176), N2=0.117 (0.087, 0.164), N0-N1).

The 16α -hydroxypregnenolone/pregnenolone ratio reflecting the 16α -hydroxylation of pregnenolone was also elevated in IgAN patients (Fig. 2C). However,

this ratio decreased after GC-application ($F=7.82$, $p=0.002$: N0=0.479 (0.393, 0.583), N1=0.366 (0.3, 0.447), N2=0.279 (0.228, 0.34), N0-N2).

Discussion

Reduced activity of adrenal zona reticularis (ZR) in IgAN patients

DHEA is an endogenous antiglucocorticoid serving as a precursor for the synthesis of 30-50 % of androgens in men and 100 % of estrogens in postmenopausal women (Labrie *et al.* 1998). At physiological concentrations, this steroid stimulates the interleukin 2 (IL-2) secretion from T helper cells type 1 (Th1) cells but suppresses tumor necrosis factor α (TNF α) and interleukin β (IL1 β) production and activation of nuclear transcription factor κ B (NF- κ B) pathway. DHEA mediates a cytotoxicity induced by T lymphocytes and enhances the natural killer (NK) cell cytotoxicity by stimulation of insulin-like growth factor 1 (IGF-1) production in NK cells. Furthermore, DHEA suppresses type 1 11 β -hydroxysteroid dehydrogenase (HSD11B1) mRNA in adipose tissue and promotes the mRNA expression of type 2 11 β -hydroxysteroid dehydrogenase (HSD11B2) in renal cells. HSD11B1 metabolite androstenediol, but not the DHEA, protects from viral-induced mortality (Loria 2002). In DHEA, DHEAS, androstenediol and DHEA/pregnenolone ratio, the between-group differences in adrenal androgens showed higher levels in the H-group in comparison with the N0-group, which indicates primarily reduced activity of the adrenal *zona reticularis* (ZR) in IgAN patients.

Glucocorticoid therapy additionally suppresses adrenal steroidogenesis in IgAN patients including the production of androgens and their immunoprotective 7- and 16 α -oxygenated metabolites

The suppression of the hormonal production in *zona fasciculata* (ZF) by prednisolone therapy was more prominent than the one in ZR as the DHEA/pregnenolone and DHEAS/pregnenolone ratios descended during the prednisolone treatment of IgAN patients. Endogenous GCs but not their synthetic analogues rapidly and dose-dependently stimulated DHEA secretion. Cortisol had no influence on steroid C17-hydroxylase-C17,20-lyase (CYP17A1) but suppressed the activity HSD3B2 activity (Topor *et al.* 2011). The lack of natural GCs in IgAN patients elevated HSD3B2 activity, which promoted the conversion of adrenal C19- Δ^5 -steroids to

their Δ^4 -counterparts and consequently reduced the levels of substrates for the synthesis of immunoprotective C19- Δ^5 -7 α / β - and 16 α -hydroxy-steroids, the synthesis of which is catalyzed by cytochrome P450 7B1 (CYP7B1), HSD11B1, cytochrome P450 3A4 (CYP3A4) and cytochrome P450 3A7 (CYP3A7). Significant or borderline decrease during the GC-treatment was obvious in all C19- Δ^5 -steroids.

Increased estradiol levels in IgAN patients irrespectively of GC-therapy and amplified conversion of DHEA to androstenedione in IgAN patients

Adrenal androgens may be peripherally converted to autoimmunity-inducing estradiol (Ahlem *et al.* 2011a, Ahlem *et al.* 2011b, Auci *et al.* 2009, Conrad *et al.* 2010, Hennebert *et al.* 2007, Le Mee *et al.* 2008, Loria 2002, Pettersson *et al.* 2010, Reading *et al.* 2012, Tang *et al.* 2006). However, in spite of reduced synthesis of C19- Δ^5 -steroids in IgAN patients we surprisingly found higher estradiol levels (independently of GC therapy) when compared with non-IgAN groups (Tables 1 and 2). The estrogen overproduction in IgAN patients may be associated with an amplified conversion of DHEA to androstenedione (catalyzed by HSD3B2) as was observed in these subjects because the androstenedione is a key estrogen precursor (Luu-The 2013). Androstenedione/DHEA and androstenedione/DHEAS ratios mounted during the GC-therapy and were higher in the T-group when compared with the H-group. We also found a different androstenedione/DHEAS ratio in the U-group when compared with both the H-group (higher values) and the T-group (lower values). Therefore, higher estradiol levels in IgAN groups may be also associated with increased aromatase activity.

Alterations in 7 α -, 7 β -, and 16 α -hydroxylation of Δ^5 -steroids

The mechanism explaining the immunomodulatory effects of 7 α / β - Δ^5 -steroids may be associated with a competition of 7-oxygenated androstanes for the active sites on the HSD11B1, which catalyzes the conversion of inactive 11-oxo-glucocorticoids to their immunosuppressive 11 β -hydroxy-counterparts (Hennebert *et al.* 2007, Le Mee *et al.* 2008).

Whereas estradiol induces autoimmunity *via* estrogen receptors, another mechanism is linked to the catabolism of C19 estrogen precursors such as DHEA, androstenediol, and 5 α -androstane-3 β ,17 β -diol (that are

also estrogenic) to their 7-oxygenated and 16 α -hydroxylated catabolites, which cannot be further converted to bioactive estrogens (Pettersson *et al.* 2010). Androstenediol, even at low concentrations is active on both types of estrogen receptors. The AT7 β , which may be either formed by interconversion from the AT7 α or directly from androstenediol by the catalytic action of CYP3A4 and CYP3A7, is immunoprotective in spite of its low concentration and high clearance (Ahlem *et al.* 2011b). The synthetic antiinflammatory derivatives of AT7 β suppress the production of C-reactive protein interleukin 17 (IL-17), TNF α , interleukin 6 (IL-6) signaling and expression of mRNAs for IL-6 and matrix metalloproteinase in inflamed tissue but intensely stimulate the splenic regulatory T cells and suppress pro-inflammatory cytokines in the lungs (Reading *et al.* 2012). Estradiol may stimulate the catalytic CYP7B1 activity, mRNA, and human CYP7B1 reporter gene in human embryonic kidney cells HEK293 and may control the DHEA, estradiol, and androstenediol levels in human tissues (Tang *et al.* 2006). We have demonstrated that DHEA7 β , but not DHEA and DHEA7 α reduces the immunosuppressive effect of GCs on the formation of plaques in murine spleen lymphocytes (Šterzl *et al.* 1999). Therefore, the elevated conversion of the C19- Δ^5 -steroids to their 7 α - and 7 β -hydroxy-metabolites (catalyzed by CYP7B1, CYP3A4, CY3A7 and HSD11B1) in IgAN patients may represent a counter-regulatory mechanism compensating the lack of C19- Δ^5 deoxysteroids by amplified synthesis of their more efficient 7-oxygenated (autoimmunity-suppressing) metabolites. The 16 α -hydroxylation of pregnenolone (catalyzed by CYP3A4, CYP3A7, CYP7B1) is also higher in IgAN patients. The only difference is the suppression of 16 α -hydroxypregnenolone/pregnenolone ratio by GCs.

Possibilities of therapeutic co-application of Δ^5 -steroids and glucocorticoids

The C19- Δ^5 -steroids mitigate the severity of ADs (Bottasso *et al.* 2007, Du *et al.* 2001, Choi *et al.* 2008, Rontzsch *et al.* 2004, Sudo *et al.* 2001, Tan *et al.* 2009) but ADs may weaken the production of adrenal C19- Δ^5 -steroids (Bottasso *et al.* 2007, Kasperska-Zajac *et al.* 2008). DHEA controls the Th1/Th2 balance and either shifts it towards the Th1 component or attenuates the

production of both components (Choi *et al.* 2008, Romagnani *et al.* 1998). The C19- Δ^5 -steroids also suppress cell-mediated immunity and formation of autoantibodies (Choi *et al.* 2008, Pratschke *et al.* 2014, Rontzsch *et al.* 2004, Sudo *et al.* 2001, Tan *et al.* 2009). Hence, the co-application of immunosuppressive corticoids with immunoprotective Δ^5 -steroids inducing restoration of the Th1-dominated cytokine profile (Hernandez-Pando *et al.* 1998b) may be favorable for the treatment of ADs. The GC-treatment alone exhibits more than 55 % risk for adverse events (Lv *et al.* 2012) and the therapy exclusively with Δ^5 -steroids may be unsafe as well (Hernandez-Pando *et al.* 1998b). The therapeutic efficiency of androstenediol is higher in comparison with DHEA and the application of adrenal androgens is optimal in an early phase of Th1-mediated response and before the switch to the Th2-mediated stage (Hernandez-Pando *et al.* 1998a).

Conclusions

The lack of adrenal androgens in IgAN patients might be a consequence of the disease linked to a lower activity of ZR. In addition, there is a further suppression of their synthesis by the GCs. Taking into account the effects of the adrenal androgens, their 7 α , 7 β , and 16 α -hydroxy-metabolites or their stable synthetic derivatives on the early stages of antibody-mediated ADs, the therapeutic co-application of the mentioned substances could be beneficial.

The finding of higher estradiol levels in IgAN patients is important as estradiol stimulates autoimmunity. Therefore, the increased conversion of C19- Δ^5 -steroids to their more efficient 7-oxygenated catabolites may be a protective mechanism against the excessive estradiol synthesis.

Conflict of Interest

There is no conflict of interest.

Acknowledgements

Supported by the grants MEYS CR (OP RDE, Excellent research – ENDO.CZ), OPPK CZ.2.16/3.1.00/21518 "Equipment for metabolomics research", and MH CZ – DRO (Institute of Endocrinology – EÚ, 00023761).

References

- AHLEM CN, AUCI DL, NICOLETTI F, PIETERS R, KENNEDY MR, PAGE TM, READING CL, ENIOUTINA EY, FRINCKE JM: Pharmacology and immune modulating properties of 5-androstene-3beta,7beta,17beta-triol, a DHEA metabolite in the human metabolome. *J Steroid Biochem Mol Biol* **126**: 87-94, 2011a.
- AHLEM CN, PAGE TM, AUCI DL, KENNEDY MR, MANGANO K, NICOLETTI F, GE Y, HUANG Y, WHITE SK, VILLEGRAS S, CONRAD D, WANG A, READING CL, FRINCKE JM: Novel components of the human metabolome: the identification, characterization and anti-inflammatory activity of two 5-androstene tetrols. *Steroids* **76**: 145-155, 2011b.
- AUCI DL, READING CL, FRINCKE JM: 7-Hydroxy androstene steroids and a novel synthetic analogue with reduced side effects as a potential agent to treat autoimmune diseases. *Autoimmun Rev* **8**: 369-372, 2009.
- BOTTASSO O, BAY ML, BESEDOVSKY H, DEL REY A: The immuno-endocrine component in the pathogenesis of tuberculosis. *Scand J Immunol* **66**: 166-175, 2007.
- CHOI IS, CUI Y, KOH YA, LEE HC, CHO YB, WON YH: Effects of dehydroepiandrosterone on Th2 cytokine production in peripheral blood mononuclear cells from asthmatics. *Korean J Intern Med* **23**: 176-181, 2008.
- CONRAD D, WANG A, PIETERS R, NICOLETTI F, MANGANO K, VAN HEECKEREN AM, WHITE SK, FRINCKE JM, READING CL, STICKNEY D, AUCI DL: HE3286, an oral synthetic steroid, treats lung inflammation in mice without immune suppression. *J Inflamm (Lond)* **7**: 52, 2010.
- DU C, KHALIL MW, SRIRAM S: Administration of dehydroepiandrosterone suppresses experimental allergic encephalomyelitis in SJL/J mice. *J Immunol* **167**: 7094-7101, 2001.
- DUNCAN DB: Multiple range and multiple F tests. *Biometrics* **11**: 1-42, 1955.
- HENNEBERT O, CHALBOT S, ALRAN S, MORFIN R: Dehydroepiandrosterone 7alpha-hydroxylation in human tissues: possible interference with type 1 11beta-hydroxysteroid dehydrogenase-mediated processes. *J Steroid Biochem Mol Biol* **104**: 326-333, 2007.
- HERNANDEZ-PANDO R, DE LA LUZ STREBER M, OROZCO H, ARRIAGA K, PAVON L, AL-NAKHLI SA, ROOK GA: The effects of androstanediol and dehydroepiandrosterone on the course and cytokine profile of tuberculosis in BALB/c mice. *Immunology* **95**: 234-241, 1998a.
- HERNANDEZ-PANDO R, DE LA LUZ STREBER M, OROZCO H, ARRIAGA K, PAVON L, MARTI O, LIGHTMAN SL, ROOK GA: Emergent immunoregulatory properties of combined glucocorticoid and anti-glucocorticoid steroids in a model of tuberculosis. *QJM* **91**: 755-766, 1998b.
- HILL M, PARIZEK A, KANCHEVA R, DUSKOVA M, VELIKOVA M, KRIZ L, KLIMKOVA M, PASKOVA A, ZIZKA Z, MATUCHA P, MELOUN M, STARKA L: Steroid metabolome in plasma from the umbilical artery, umbilical vein, maternal cubital vein and in amniotic fluid in normal and preterm labor. *J Steroid Biochem Mol Biol* **121**: 594-610, 2010.
- KASPERSKA-ZAJAC A, BRZOZA Z, ROGALA B: Dehydroepiandrosterone and dehydroepiandrosterone sulphate in atopic allergy and chronic urticaria. *Inflammation* **31**: 141-145, 2008.
- LABRIE F, BELANGER A, LUU-THE V, LABRIE C, SIMARD J, CUSAN L, GOMEZ JL, CANDAS B: DHEA and the intracrine formation of androgens and estrogens in peripheral target tissues: its role during aging. *Steroids* **63**: 322-328, 1998.
- LAI KN, TANG SC, SCHENA FP, NOVAK J, TOMINO Y, FOGO AB, GLASSOCK RJ: IgA nephropathy. *Nat Rev Dis Primers* **2**: 16001, 2016.
- LE MEE S, HENNEBERT O, FERREC C, WULFERT E, MORFIN R: 7beta-hydroxy-epiandrosterone-mediated regulation of the prostaglandin synthesis pathway in human peripheral blood monocytes. *Steroids* **73**: 1148-1159, 2008.
- LORIA RM: Immune up-regulation and tumor apoptosis by androstene steroids. *Steroids* **67**: 953-966, 2002.
- LUU-THE V: Assessment of steroidogenesis and steroidogenic enzyme functions. *J Steroid Biochem Mol Biol* **137**: 176-182, 2013.
- LV J, XU D, PERKOVIC V, MA X, JOHNSON DW, WOODWARD M, LEVIN A, ZHANG H, WANG H: Corticosteroid therapy in IgA nephropathy. *J Am Soc Nephrol* **23**: 1108-1116, 2012.

- MASTORAKOS G, KAROUTSOU EI, MIZAMTSIDI M: Corticotropin releasing hormone and the immune/inflammatory response. *Eur J Endocrinol* **155**: S77-S84, 2006.
- MATOUSOVIC K, MESTECKY J, VONDRAK K, DUSEK J, CHVATALOVA E, HACEK J, HORYNOVA M, KASPEROVA A, ROSSMANN P, STERZL I, RASKA M: IgA Nephropathy. Facts, uncertainties, potential causal therapy approaches (in Czech). *Cas Lek Cesk* **154**: 168-173, 2015.
- MESTECKY J, RASKA M, JULIAN BA, GHARAVI AG, RENFROW MB, MOLDOVEANU Z, NOVAK L, MATOUSOVIC K, NOVAK J: IgA nephropathy: molecular mechanisms of the disease. *Annu Rev Pathol* **8**: 217-240, 2013.
- PETTERSSON H, LUNDQVIST J, NORLIN M: Effects of CYP7B1-mediated catalysis on estrogen receptor activation. *Biochim Biophys Acta* **1801**: 1090-1097, 2010.
- PRATSCHKE S, VON DOSSOW-HANFSTINGL V, DIETZ J, SCHNEIDER CP, TUFMAN A, ALBERTSMEIER M, WINTER H, ANGELE MK: Dehydroepiandrosterone modulates T-cell response after major abdominal surgery. *J Surg Res* **189**: 117-125, 2014.
- RAUEN T, EITNER F, FITZNER C, SOMMERER C, ZEIER M, OTTE B, PANZER U, PETERS H, BENCK U, MERTENS PR, KUHLMANN U, WITZKE O, GROSS O, VIELHAUER V, MANN JF, HILGERS RD, FLOEGE J: Intensive supportive care plus immunosuppression in IgA nephropathy. *N Engl J Med* **373**: 2225-2236, 2015.
- READING CL, FRINCKE JM, WHITE SK: Molecular targets for 17alpha-ethynodiol-5-androstene-3beta,7beta,17beta-triol, an anti-inflammatory agent derived from the human metabolome. *PLoS One* **7**: e32147, 2012.
- RENGARAJAN S, BALASUBRAMANIAN K: Corticosterone induces steroidogenic lesion in cultured adult rat Leydig cells by reducing the expression of star protein and steroidogenic enzymes. *J Cell Biochem* **103**: 1472-1487, 2008.
- ROMAGNANI S, KAPSENBERG M, RADBRUCH A, ADORINI L: Th1 and Th2 cells. *Res Immunol* **149**: 871-873, 1998.
- RONTZSCH A, THOSS K, PETROW PK, HENZGEN S, BRAUER R: Amelioration of murine antigen-induced arthritis by dehydroepiandrosterone (DHEA). *Inflamm Res* **53**: 189-198, 2004.
- STERZL I, HAMPL R, STERZL J, VOTRUBA J, STARKA L: 7Beta-OH-DHEA counteracts dexamethasone induced suppression of primary immune response in murine spleenocytes. *J Steroid Biochem Mol Biol* **71**: 133-137, 1999.
- SUDO N, YU XN, KUBO C: Dehydroepiandrosterone attenuates the spontaneous elevation of serum IgE level in NC/Nga mice. *Immunol Lett* **79**: 177-179, 2001.
- TAN XD, DOU YC, SHI CW, DUAN RS, SUN RP: Administration of dehydroepiandrosterone ameliorates experimental autoimmune neuritis in Lewis rats. *J Neuroimmunol* **207**: 39-44, 2009.
- TANG W, EGGERTSEN G, CHIANG JY, NORLIN M: Estrogen-mediated regulation of CYP7B1: a possible role for controlling DHEA levels in human tissues. *J Steroid Biochem Mol Biol* **100**: 42-51, 2006.
- TESAR V, TROYANOV S, BELLUR S, VERHAVE JC, COOK HT, FEEHALY J, ROBERTS IS, CATTRAN D, COPPO R: Corticosteroids in IgA nephropathy: a retrospective analysis from the VALIGA study. *J Am Soc Nephrol* **26**: 2248-2258, 2015.
- TOPOR LS, ASAI M, DUNN J, MAJZOUB JA: Cortisol stimulates secretion of dehydroepiandrosterone in human adrenocortical cells through inhibition of 3betaHSD2. *J Clin Endocrinol Metab* **96**: E31-E39, 2011.